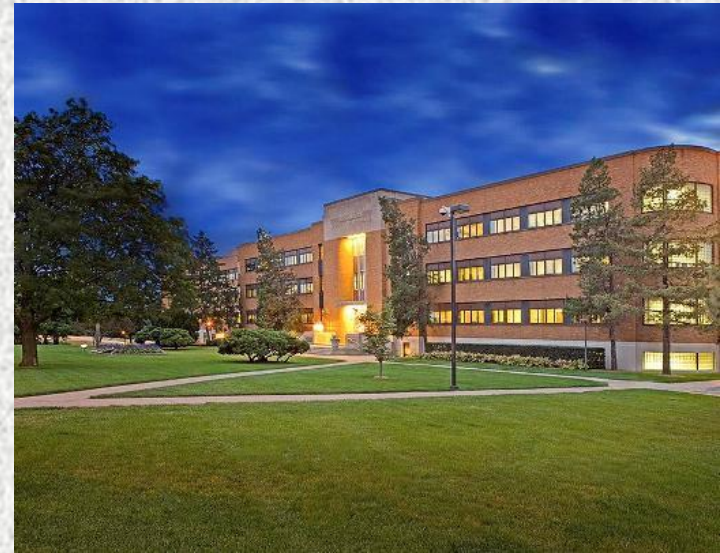


Biological Abatement of Fermentation Inhibitors

Nancy N. Nichols, Badal C. Saha, Bruce S. Dien, Michael A. Cotta

National Center for Agricultural Utilization Research
ARS-USDA, Peoria, IL



Biomass to ethanol

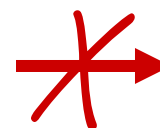


Cellulose
Hemicellulose



Enzymes
Acid

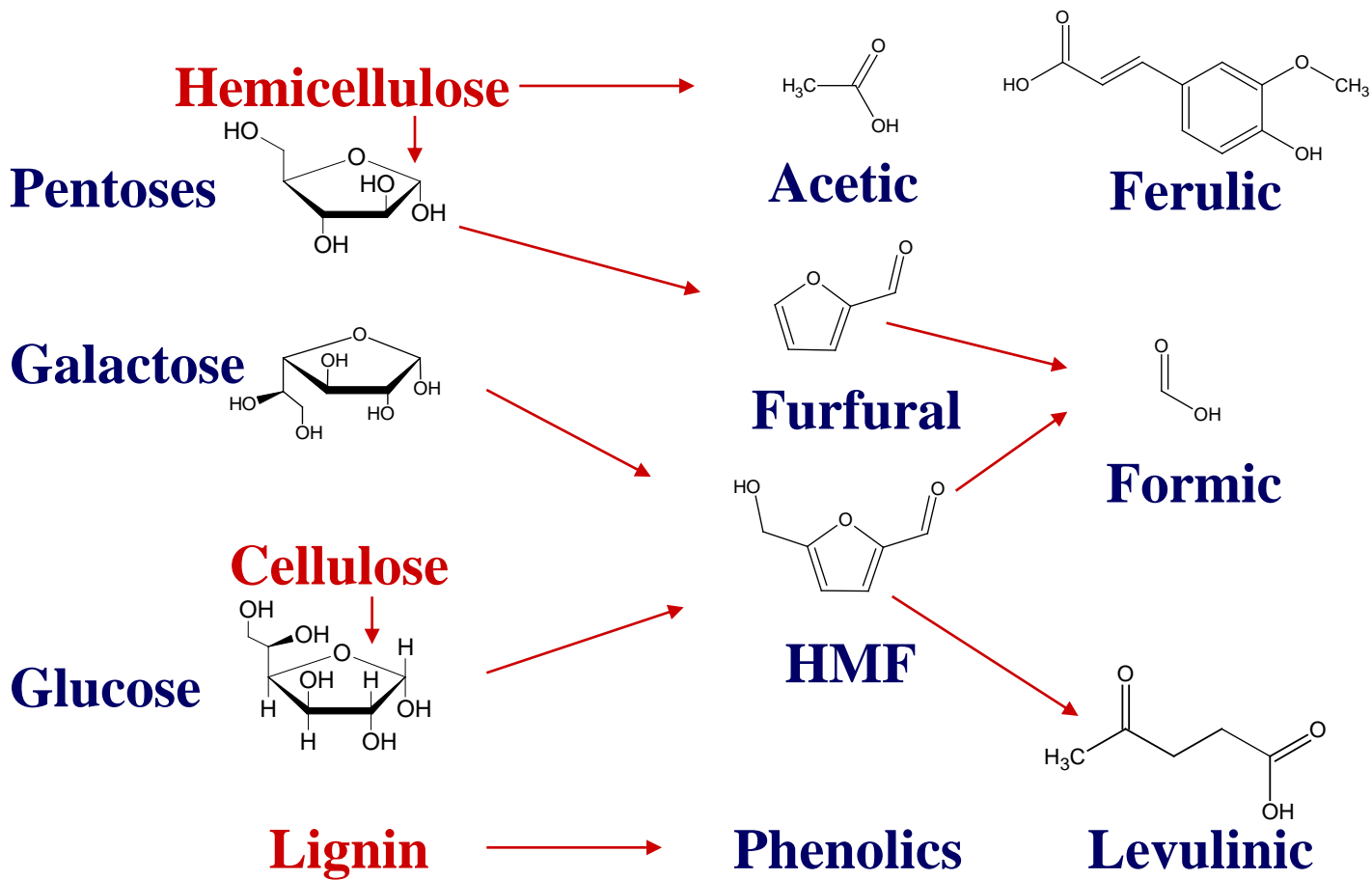
Hexoses
Pentoses
Inhibitors



Recombinant
Yeast or Bacteria

Ethanol

Inhibitors formed during hydrolysis

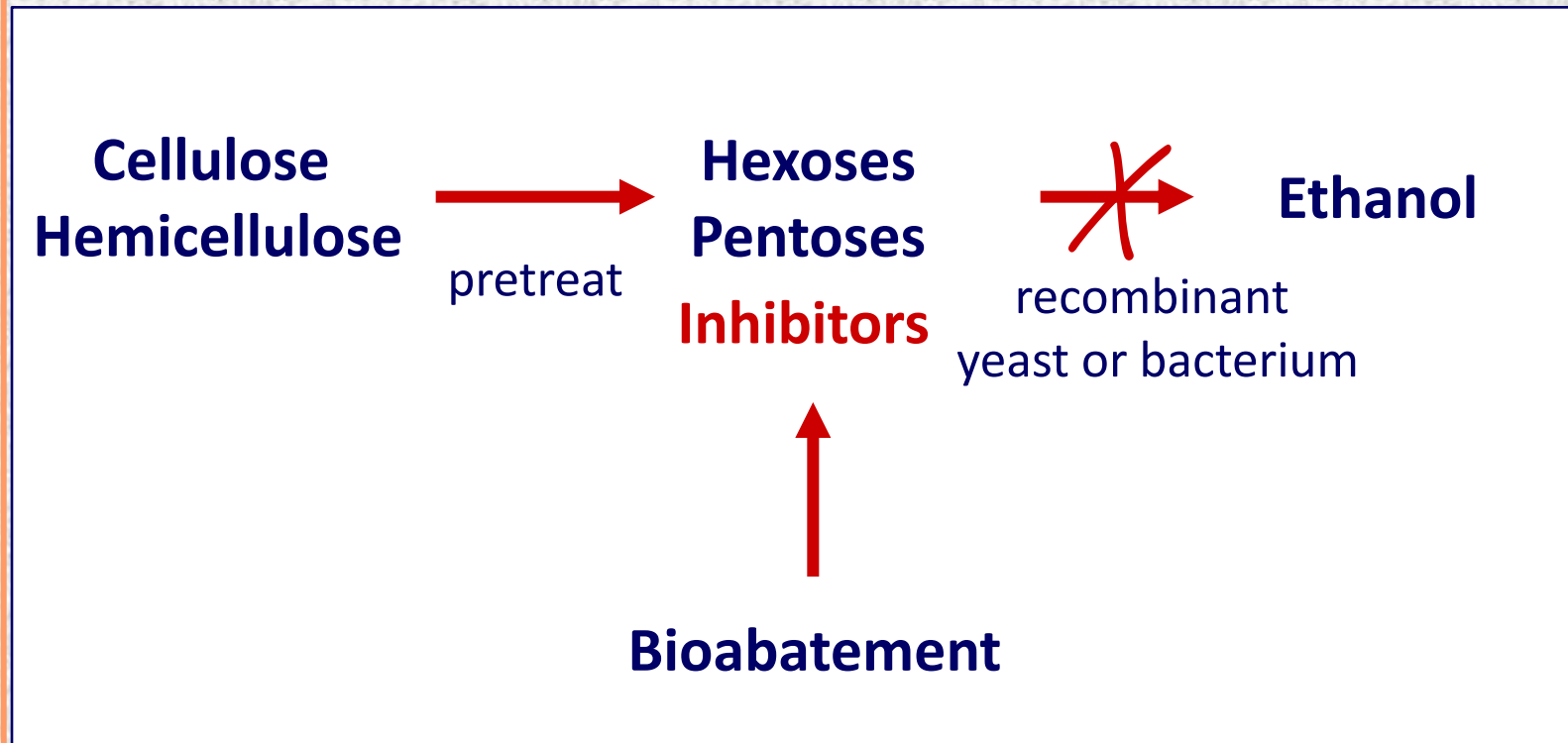


Inhibitor abatement



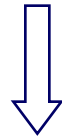
- ◆ Inexpensive
- ◆ Easy to integrate
- ◆ Selective removal of inhibitors
- ◆ Dilution
 - Vacuum evaporation
 - pH adjustment
 - Polymeric adsorption

A microbiological approach to inhibitor abatement

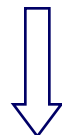


Enrichment of detoxifying microbes

Environmental sample: contaminated soil



Enrichment in defined media

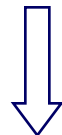


- Furfural
- 5-HMF
- Ferulic acid



Microorganisms
metabolize
toxic compounds

Enrichment in Hydrolysate



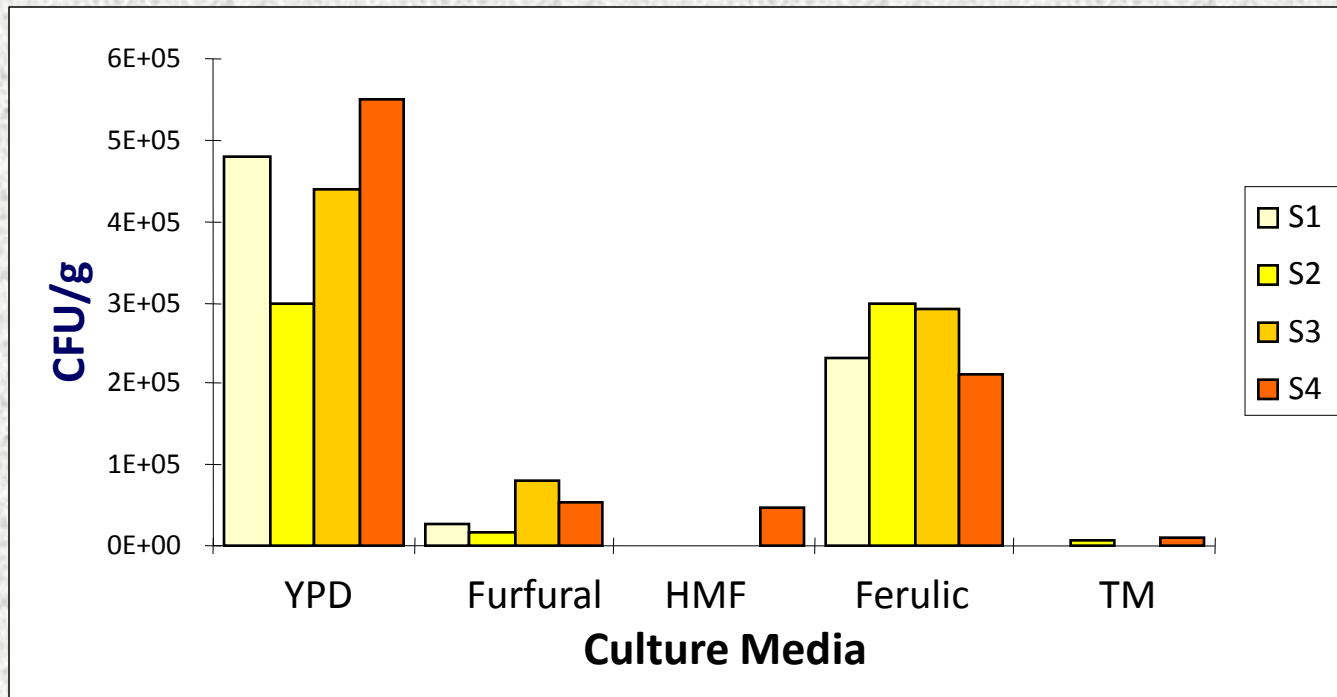
Microorganisms
survive/grow in
dilute acid
hydrolysates

Isolation and identification



Microbes enriched from contaminated soil

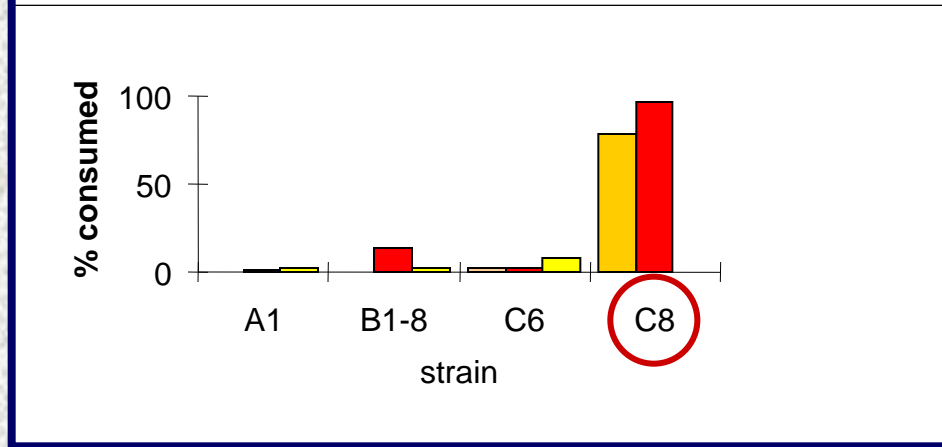
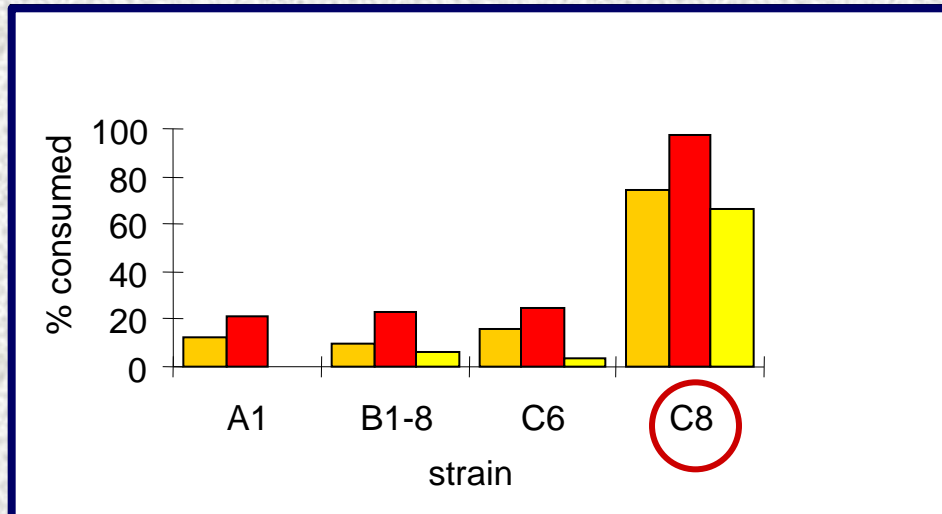
Direct Microbial Counts from Four Soil Samples



Identification of isolates

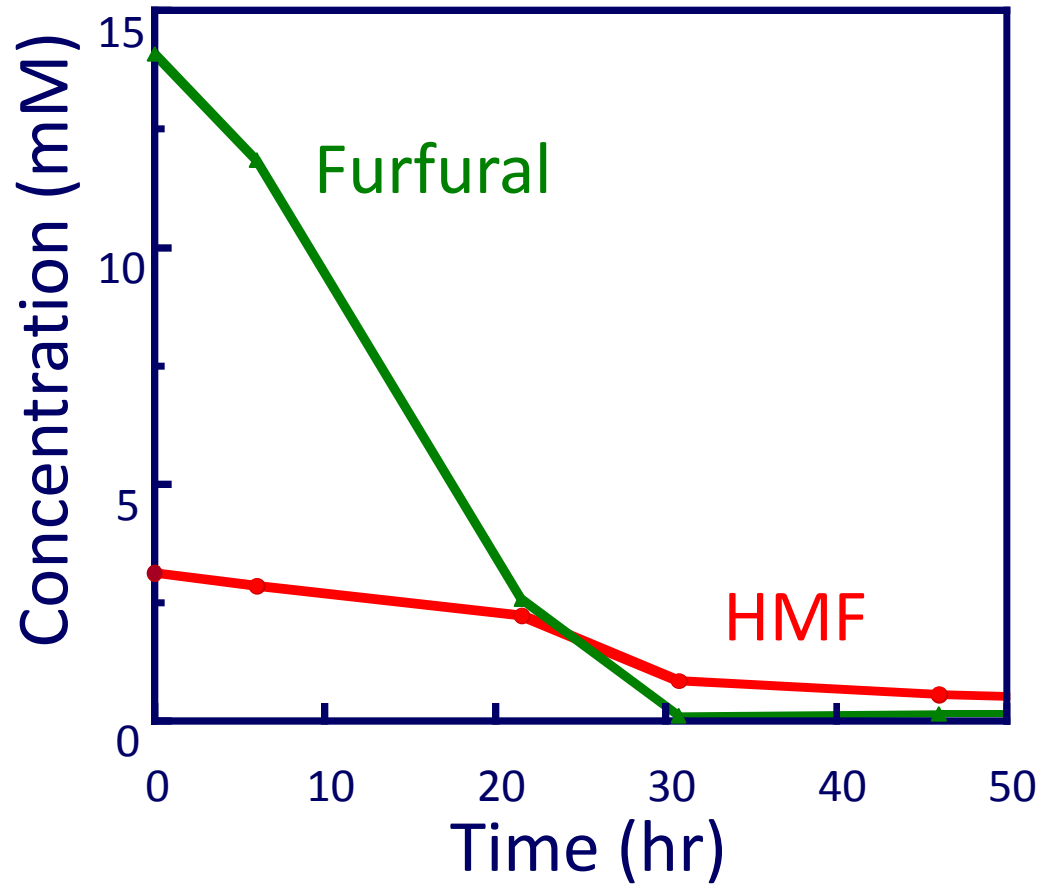
- ◆ A3, A6, B1-1: *Pseudomonas sp.*
- ◆ A1: *Methylobacterium extorquens*
- ◆ A5: *Arthrobacter aurescens*
- ◆ B1-1: *Acinetobacter sp.*
- ◆ C6: *Flavobacterium indologenes*
- ◆ C7: *Stenotrophomonas maltophilia*
- ◆ C8: Ascomycete fungus

Treatment of hydrolysate

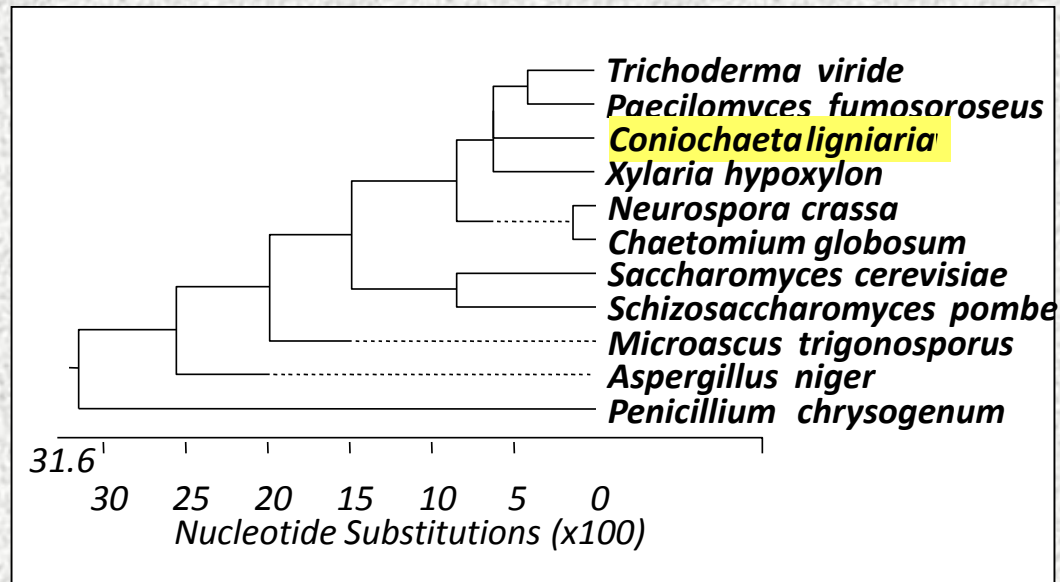


■ HMF
■ Furfural
■ Ferulic

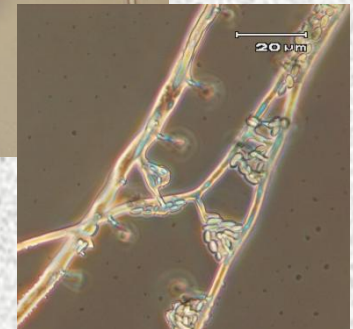
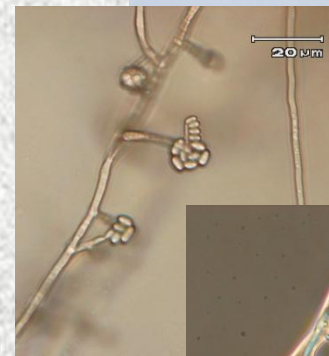
Removal of furans from corn stover hydrolysate



Coniochaeta ligniaria C8 (NRRL30616)



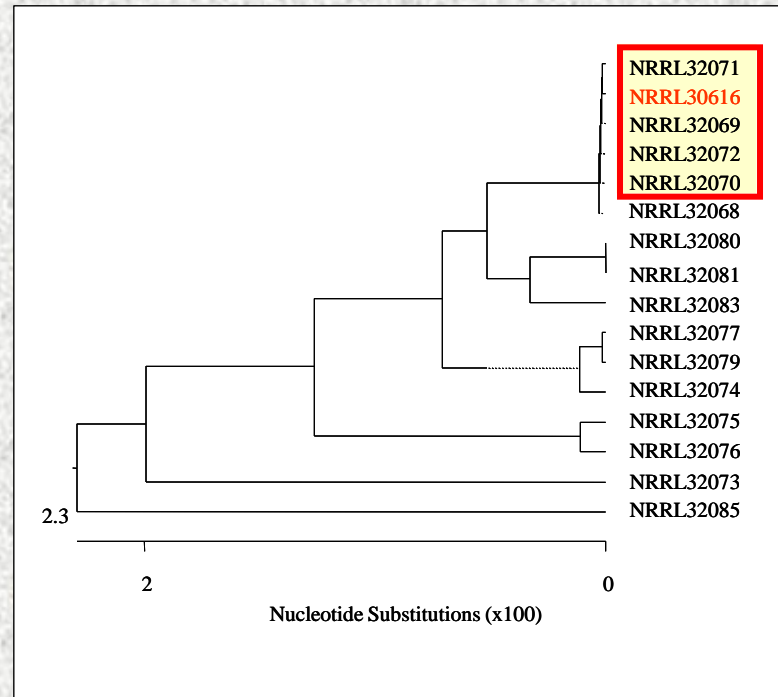
Alignment of the large ribosomal subunit (D1D2 domain) of representative Ascomycetes by the ClustalW method. (Adapted from Weber et al (2002)).



Comparison of *Coniochaeta* strains

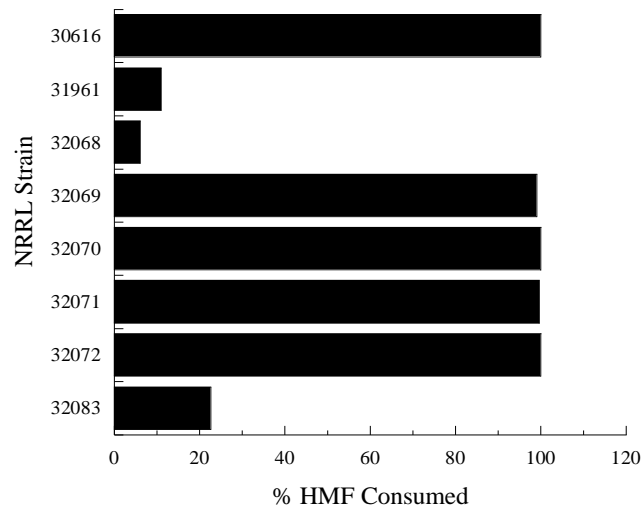
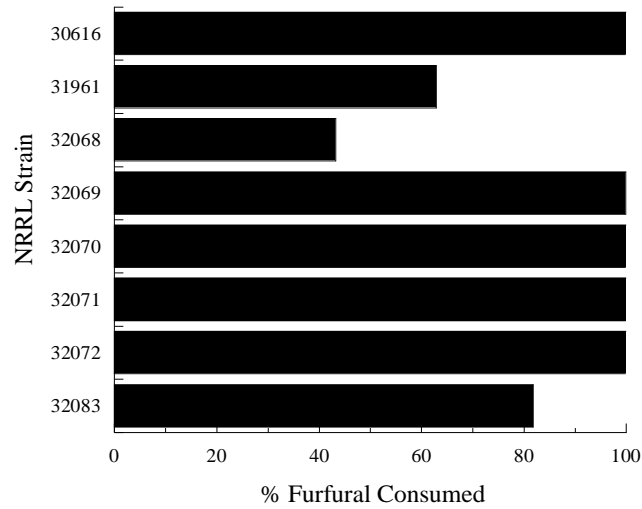
Organism	NRRL	Other ID	Furfural	5-HMF	Levulinic Acid	<i>p</i> -Hydroxybenzaldehyde
<i>Coniochaeta ligniaria</i>	30616	C8	0.605	0.090	0.006	0.081
<i>Lecytophora hoffmannii</i>	31961	DSM2693	0.560	0.148	0	0.086
<i>Lecytophora mutabilis</i>	31962	DSM10716	---	---	n.d.	---
<i>Lecytophora lignicola</i>	31963	DSM63551	---	---	n.d.	---
<i>Coniochaeta ligniaria</i>	32068	95.605	0.278	0.023	0.308	---
<i>Coniochaeta ligniaria</i>	32069	98.1105	0.577	0.166	0.145	0.063
<i>Coniochaeta ligniaria</i>	32070	98.1126	0.640	0.101	0.073	0.196
<i>Coniochaeta ligniaria</i>	32071	F3331	0.005	0.104	n.d.	0.136
<i>Coniochaeta ligniaria</i>	32072	F3343	0.723	0.214	0.148	0.073
<i>Phialophora decumbens</i>	32073	CBS153.42	---	---	n.d.	---
<i>Phialophora fasciculatus</i>	32074	CBS205.38	---	---	n.d.	---
<i>Lecytophora hoffmannii</i>	32075	CBS245.38	---	---	n.d.	---
<i>Lecytophora hoffmannii</i>	32076	CBS140.41	---	---	n.d.	0.053
<i>Lecytophora lignicola</i>	32077	CBS267.33	---	---	n.d.	---
<i>Phialophora luteoviridis</i>	32079	CBS206.38	---	---	n.d.	---
<i>Lecytophora mutabilis</i>	32080	CBS157.44	---	---	n.d.	---
<i>Lecytophora mutabilis</i>	32081	CBS303.62	---	---	n.d.	0.09
<i>Coniochaeta ligniaria</i>	32082	CBS620.69	---	---	n.d.	---
<i>Coniochaeta ligniaria</i>	32083	CBS178.75	0.186	0.071	0.088	---
<i>Coniochaeta malacotricha</i>	32084	CBS323.72	---	---	n.d.	---

Comparison of *Coniochaeta* strains

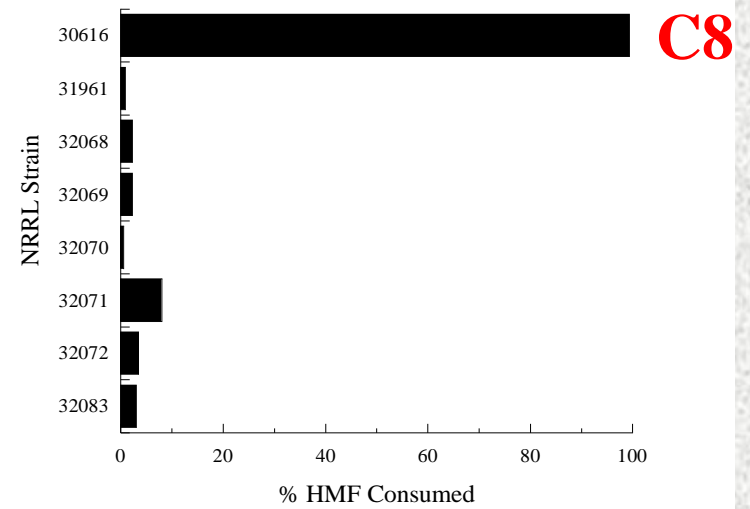
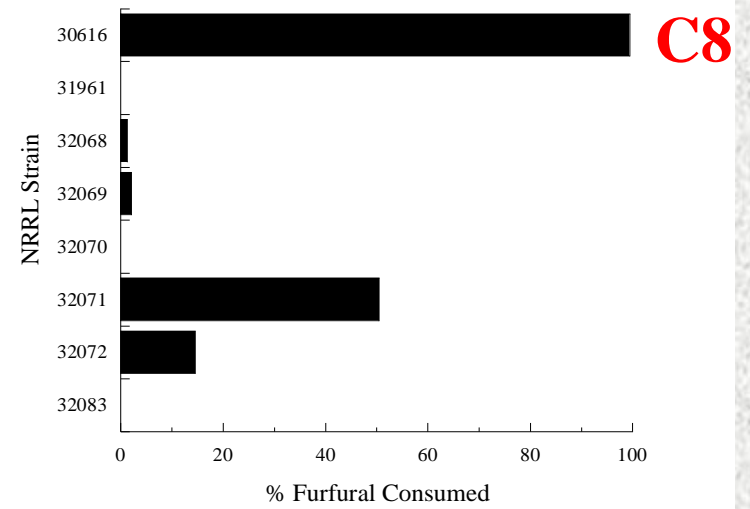


Alignment of the large ribosomal subunit (D1D2 domain) of the strains in Table 1 by the ClustalW method (Adapted from Weber et al (2002)). Strains that were the best at removing furfural from corn stover hydrolysate are highlighted.

0.75X

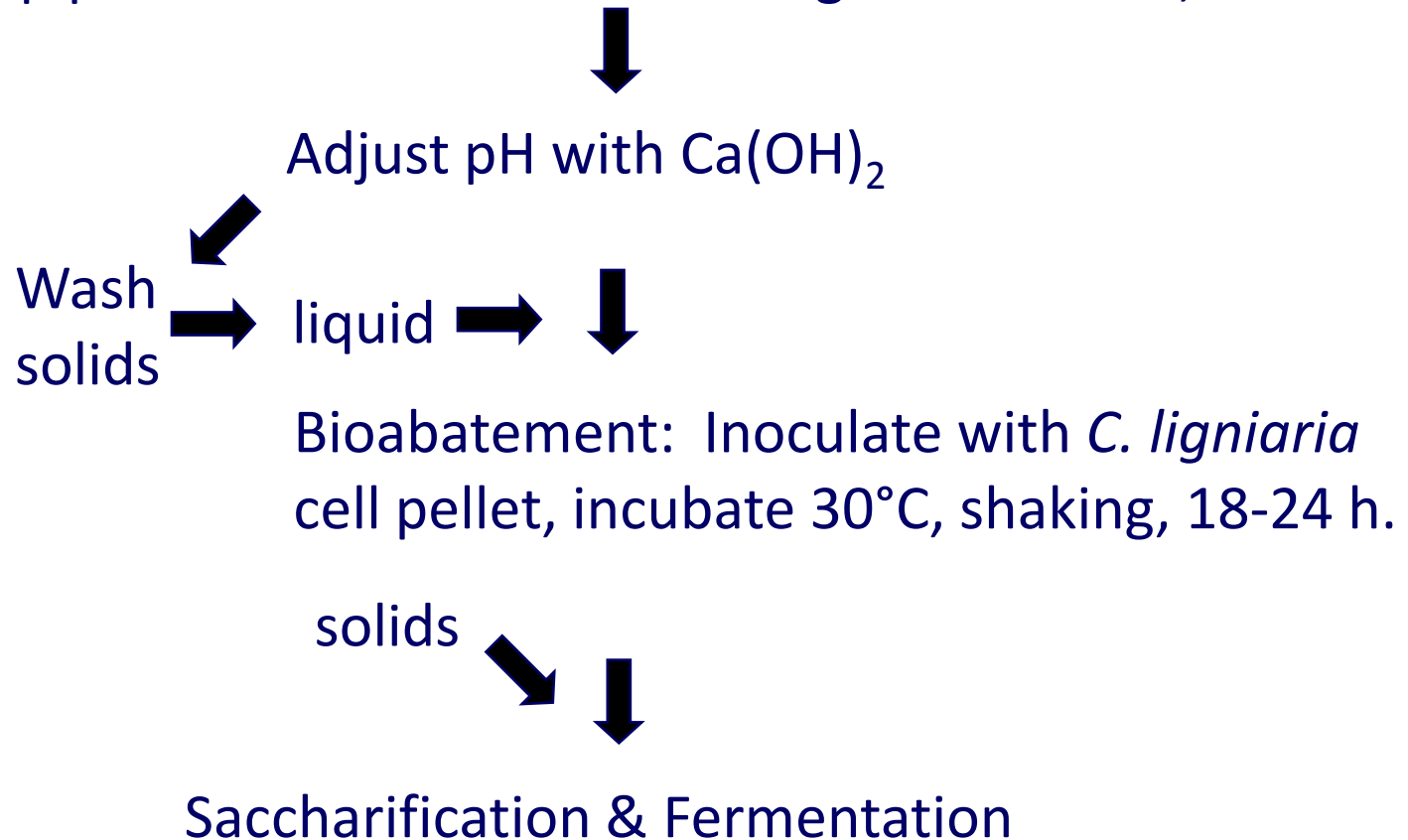


1X

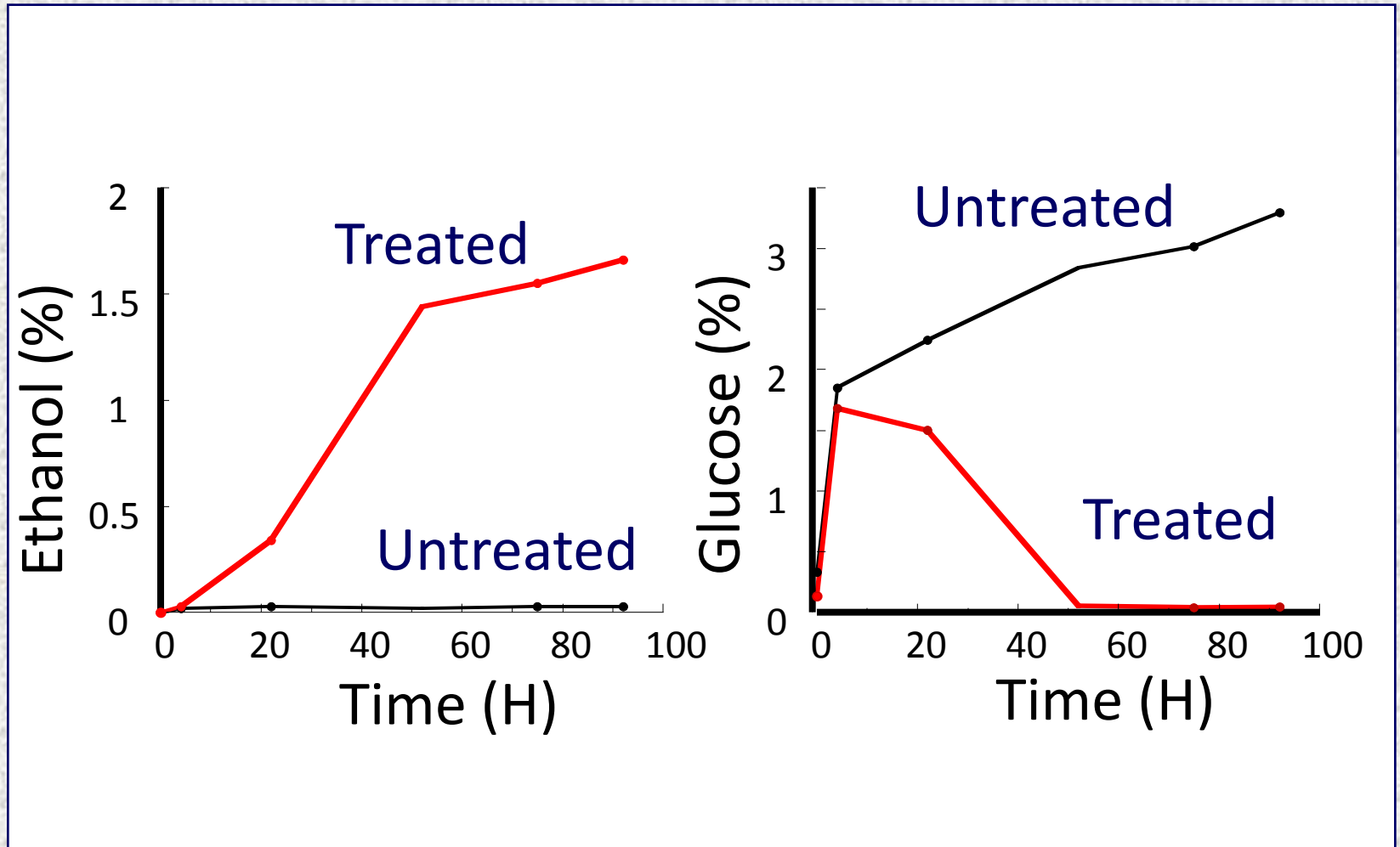


General bioabatement scheme

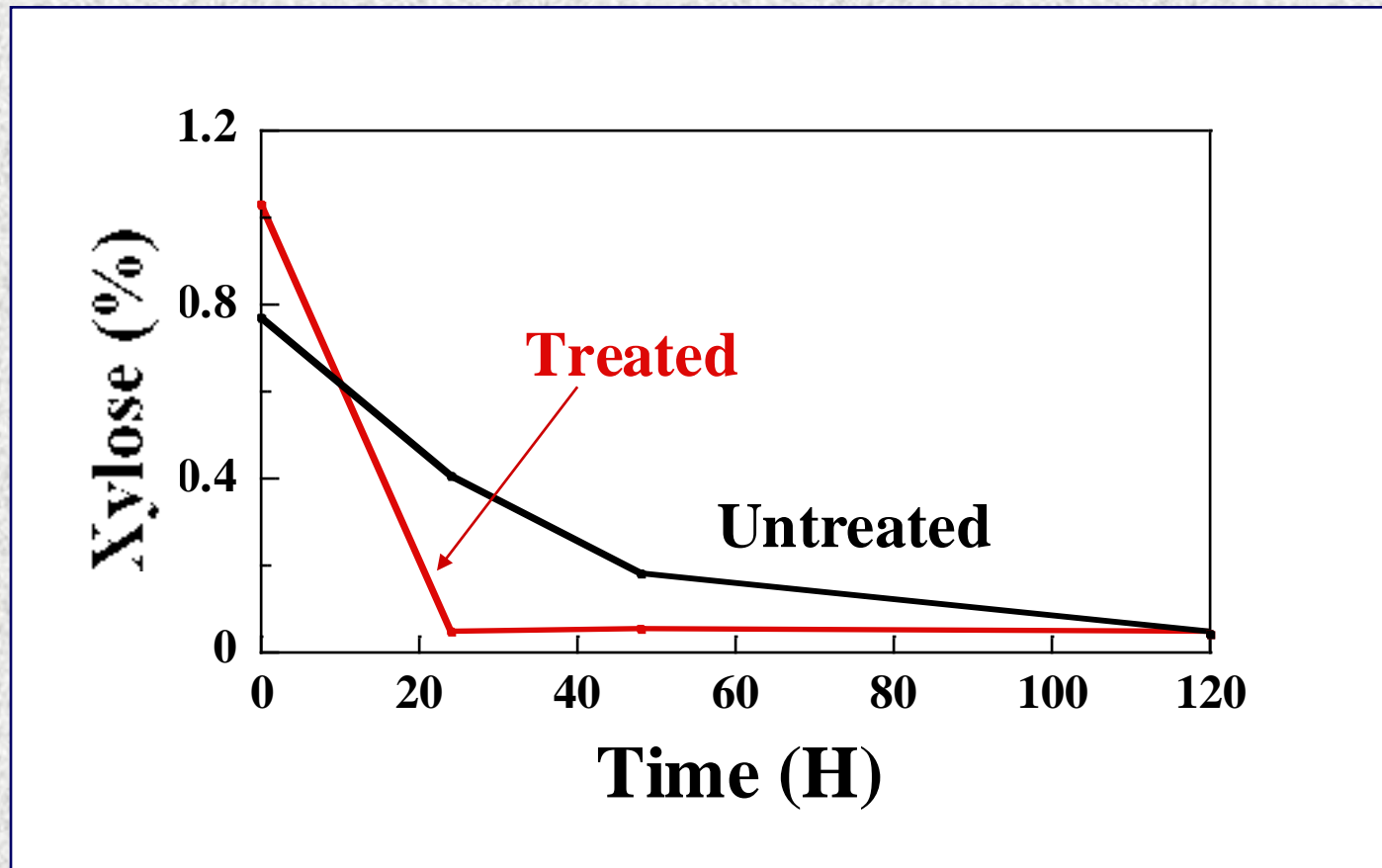
10 % corn stover and 0.66% H₂SO₄
Steel pipe reactors – fluidized heating bath –180°C, 10 min.



Fermentation of glucose in corn stover

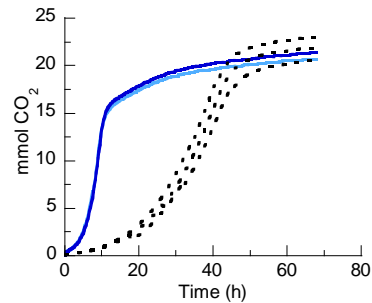


Fermentation of xylose in corn stover

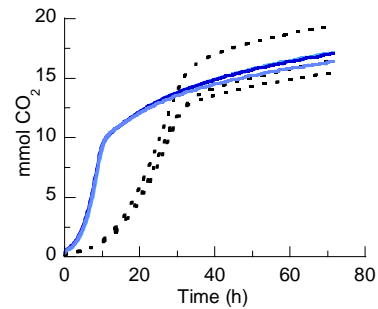


Reed Canarygrass

Vegetative

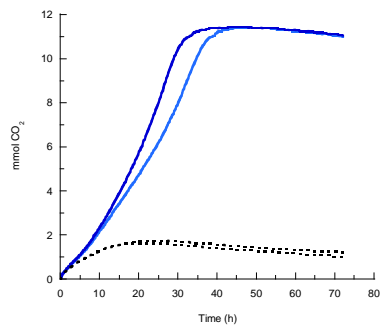


Ripe Seed

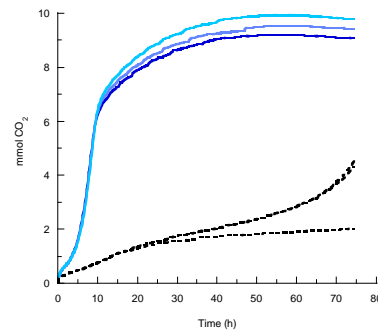


Alfalfa Stems

Bud

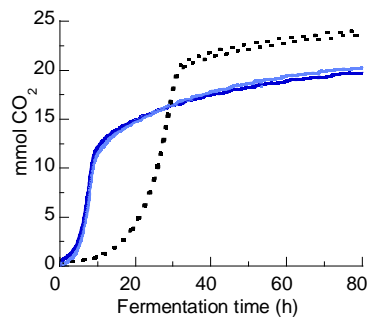


Flower

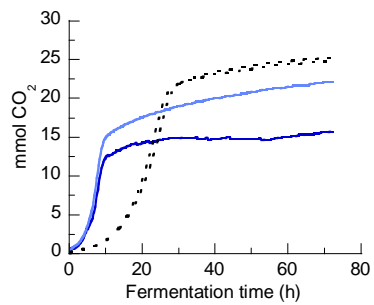


Switchgrass

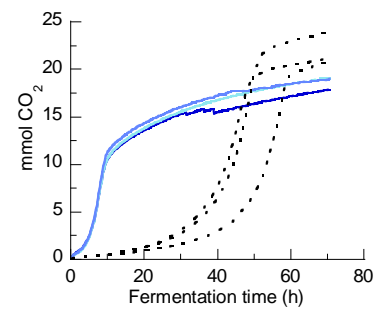
Preboot



Anthesis

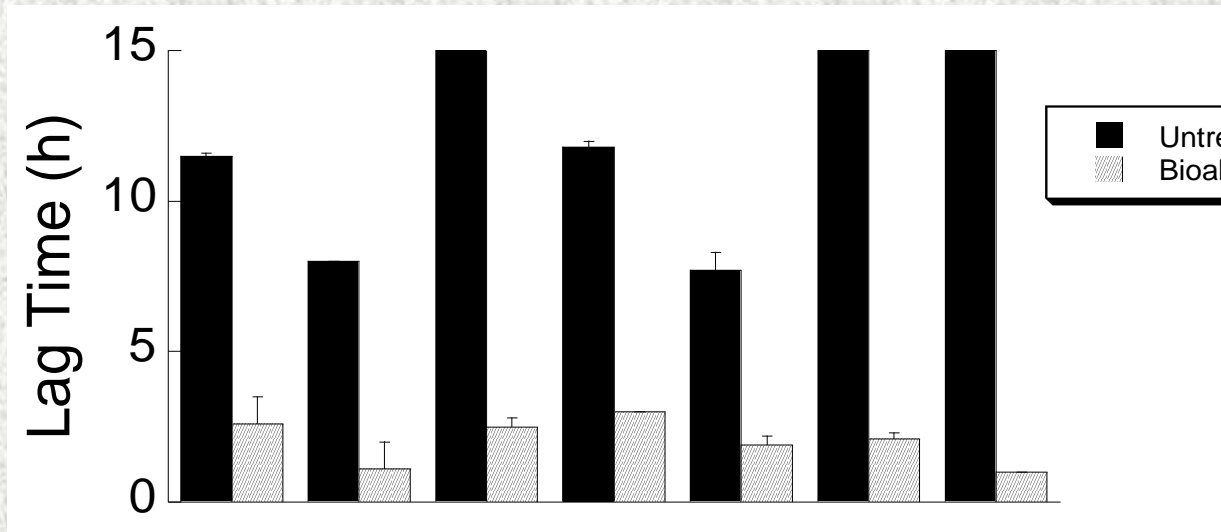


Post frost

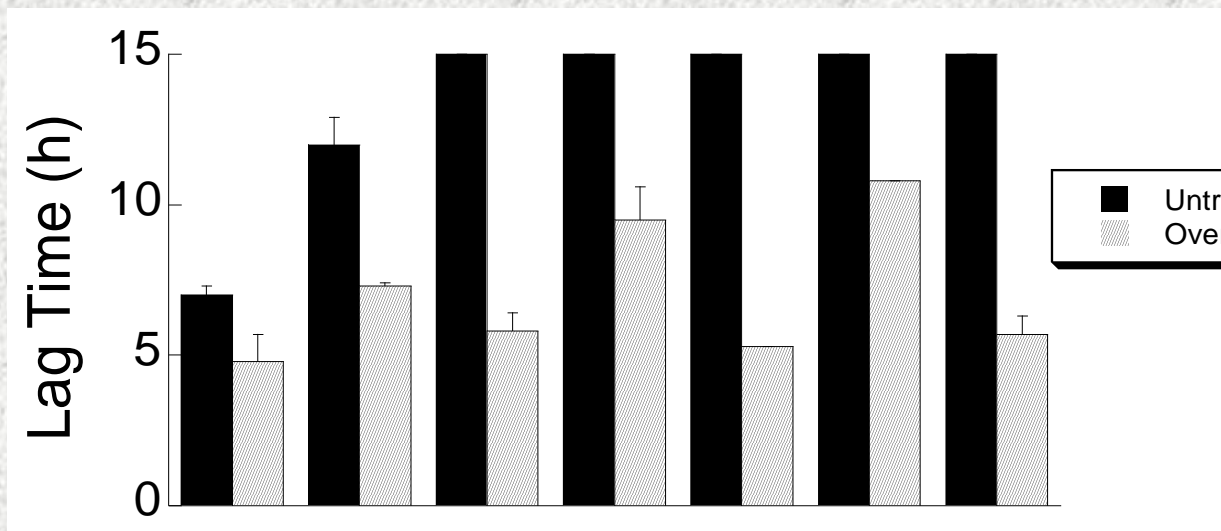


Fermentation lag time

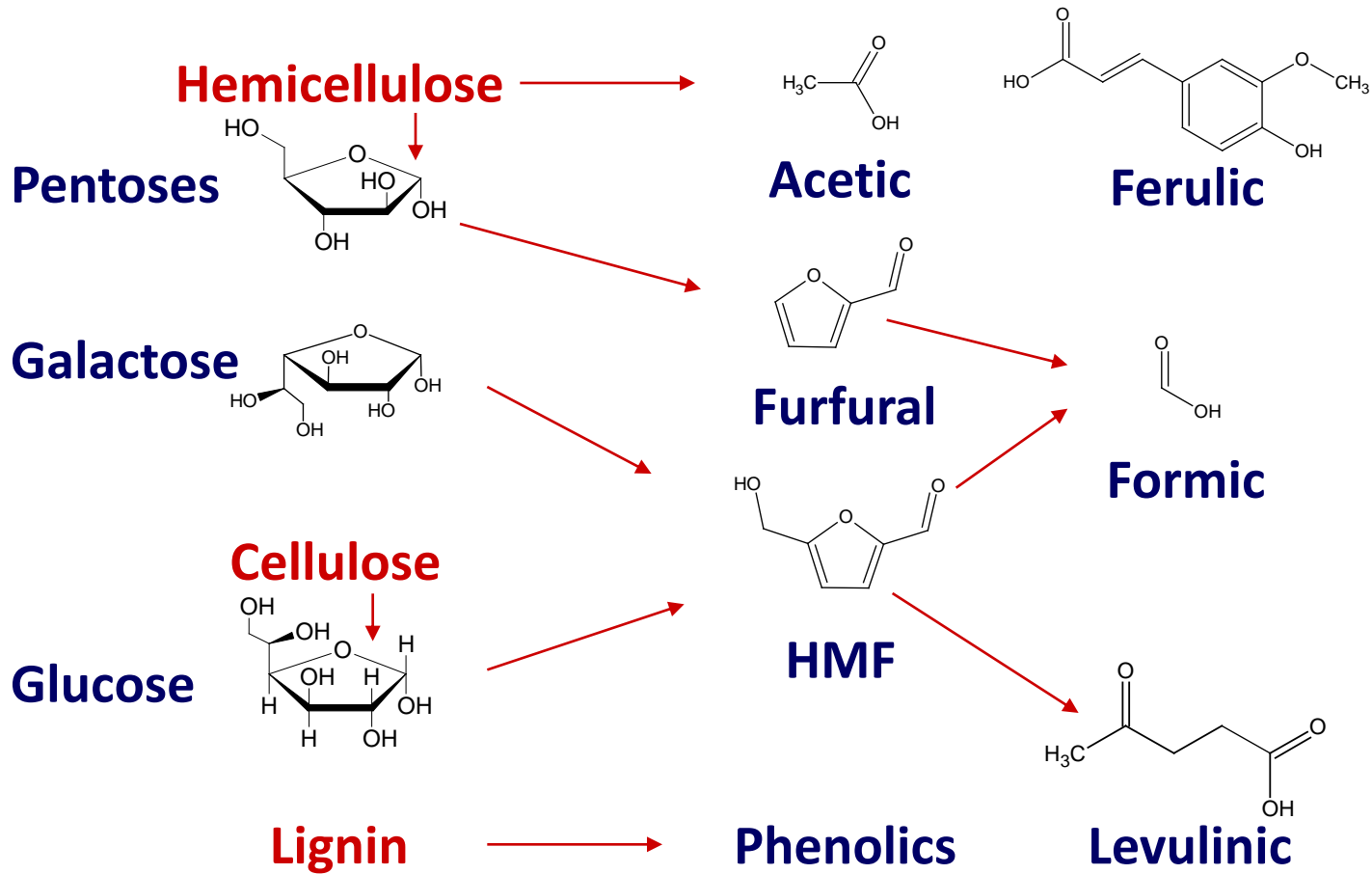
Bioabatement



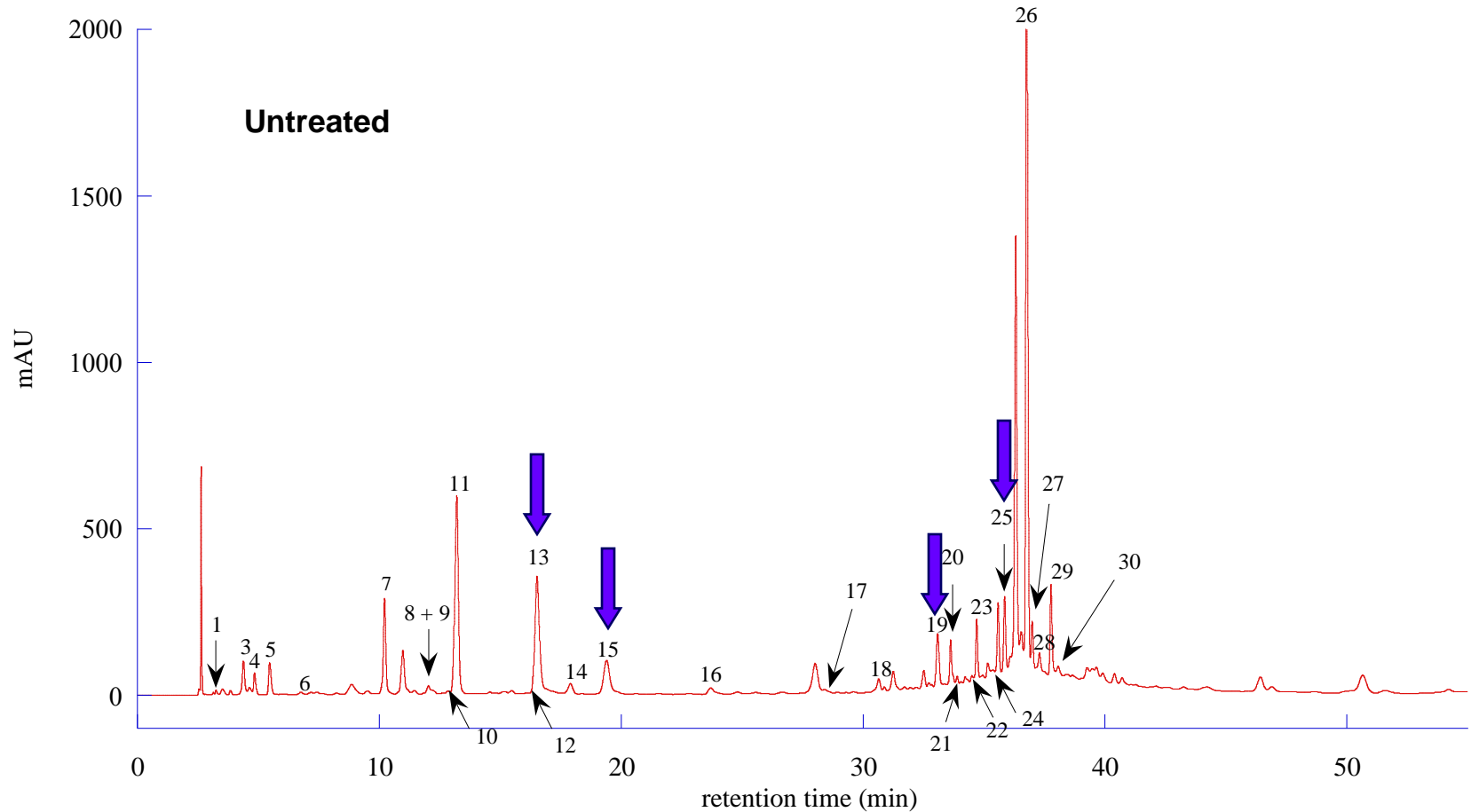
Overliming



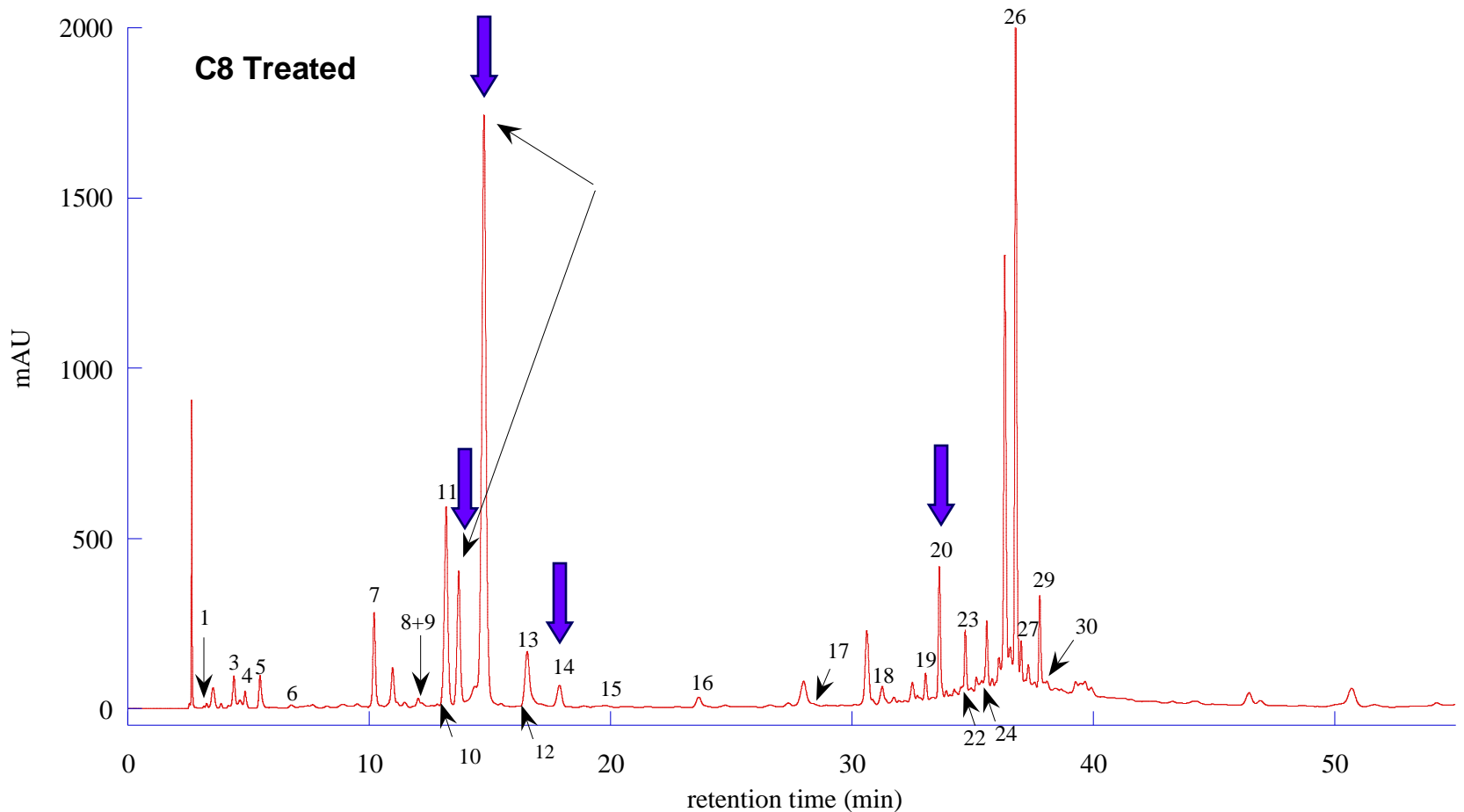
Inhibitors formed during hydrolysis



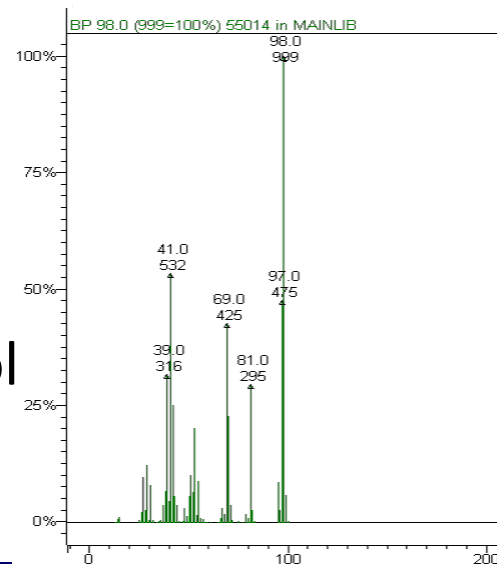
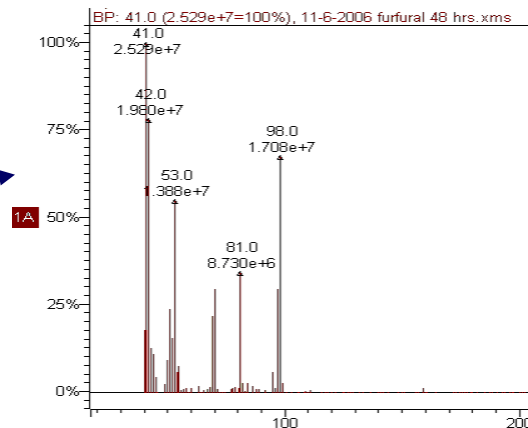
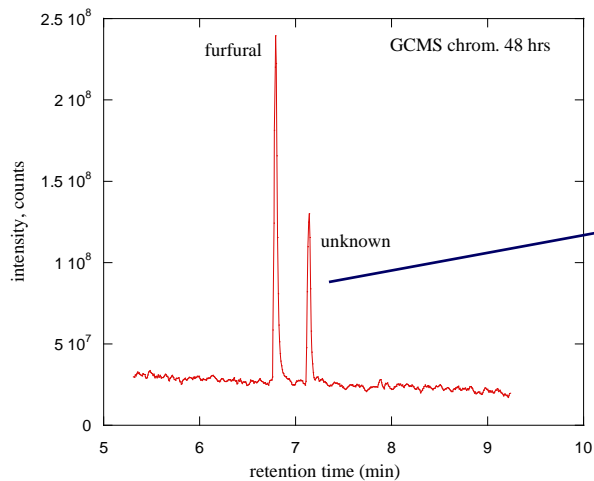
Untreated corn stover hydrolysate



C8-treated corn stover hydrolysate



New peak is furfuryl alcohol

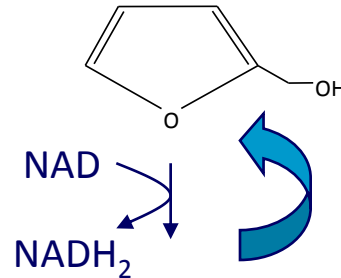


NIST
2-furanmethanol

Furan metabolic pathway

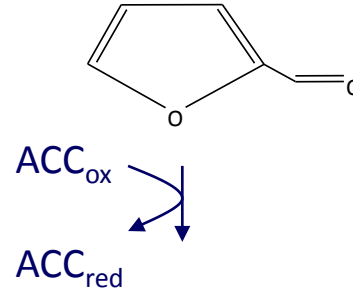
Furfuryl Alcohol

Furfuryl Alcohol
Dehydrogenase



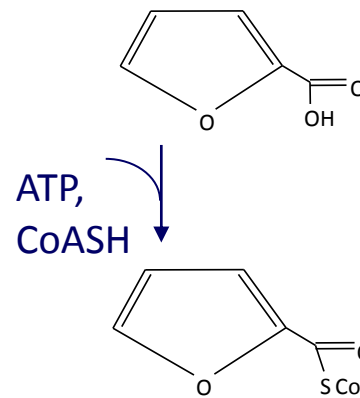
Furfural

Furfural
Dehydrogenase



2-Furoic Acid

2-Furoate-CoA
Synthetase

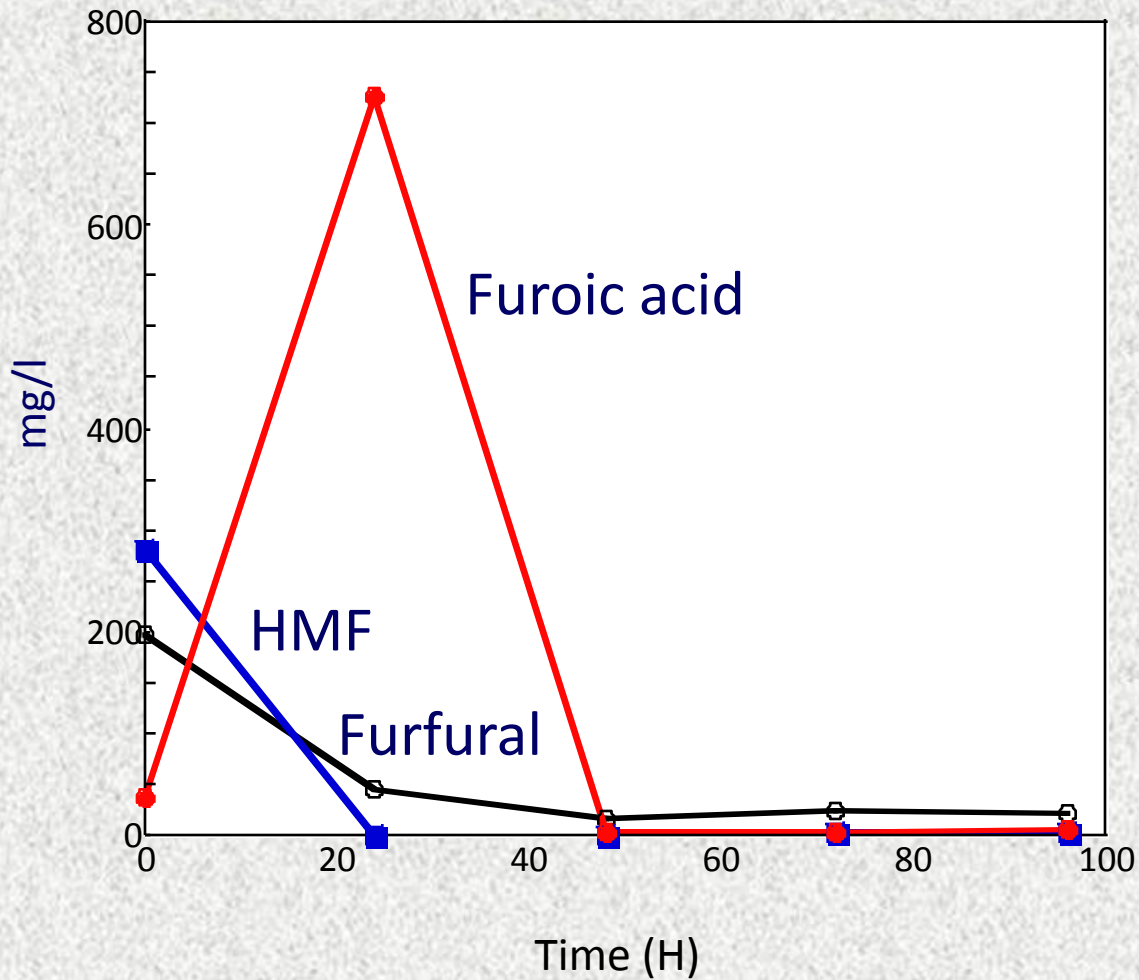


2-Furoyl-CoA

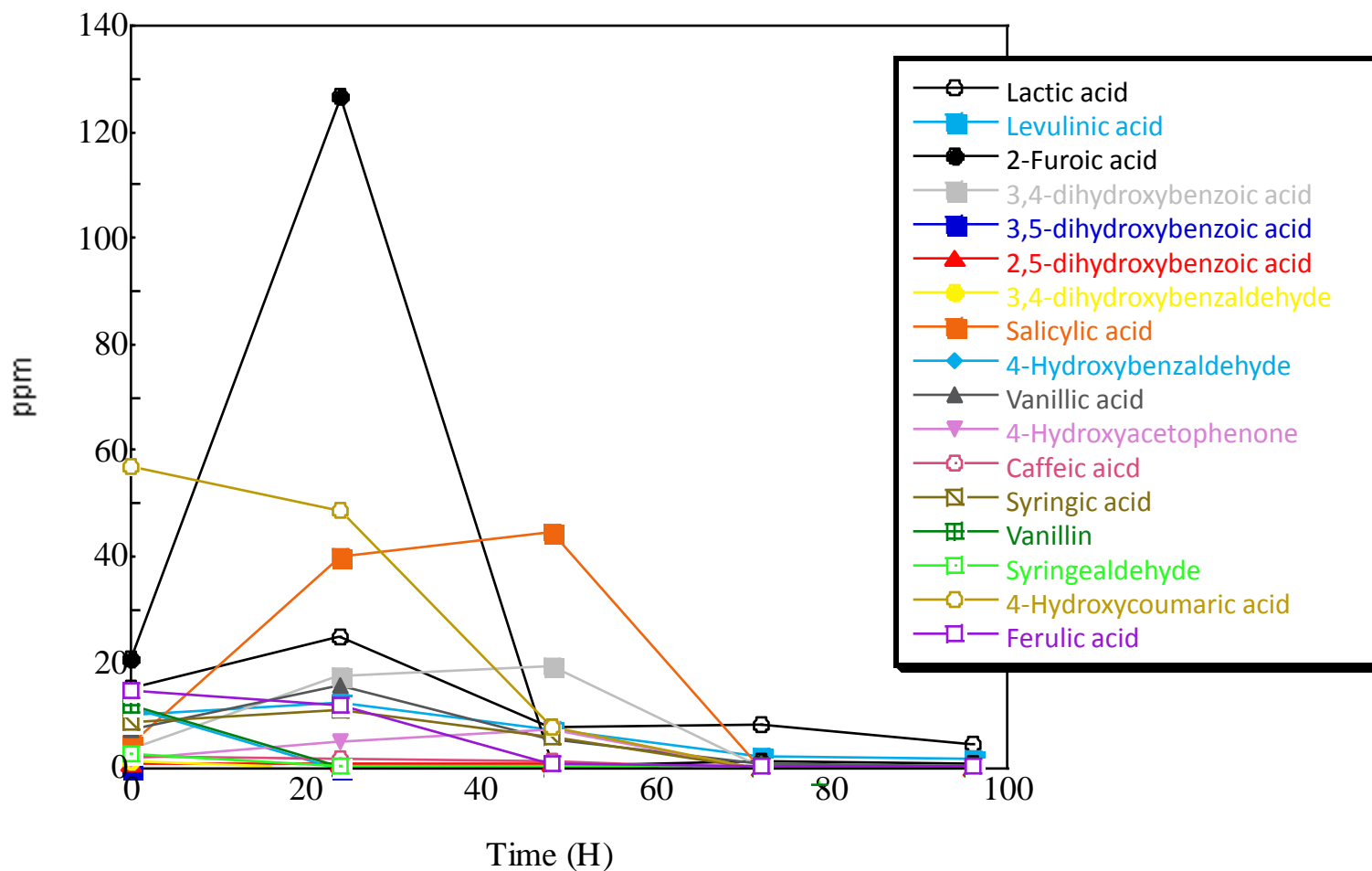


Citric Acid
Cycle

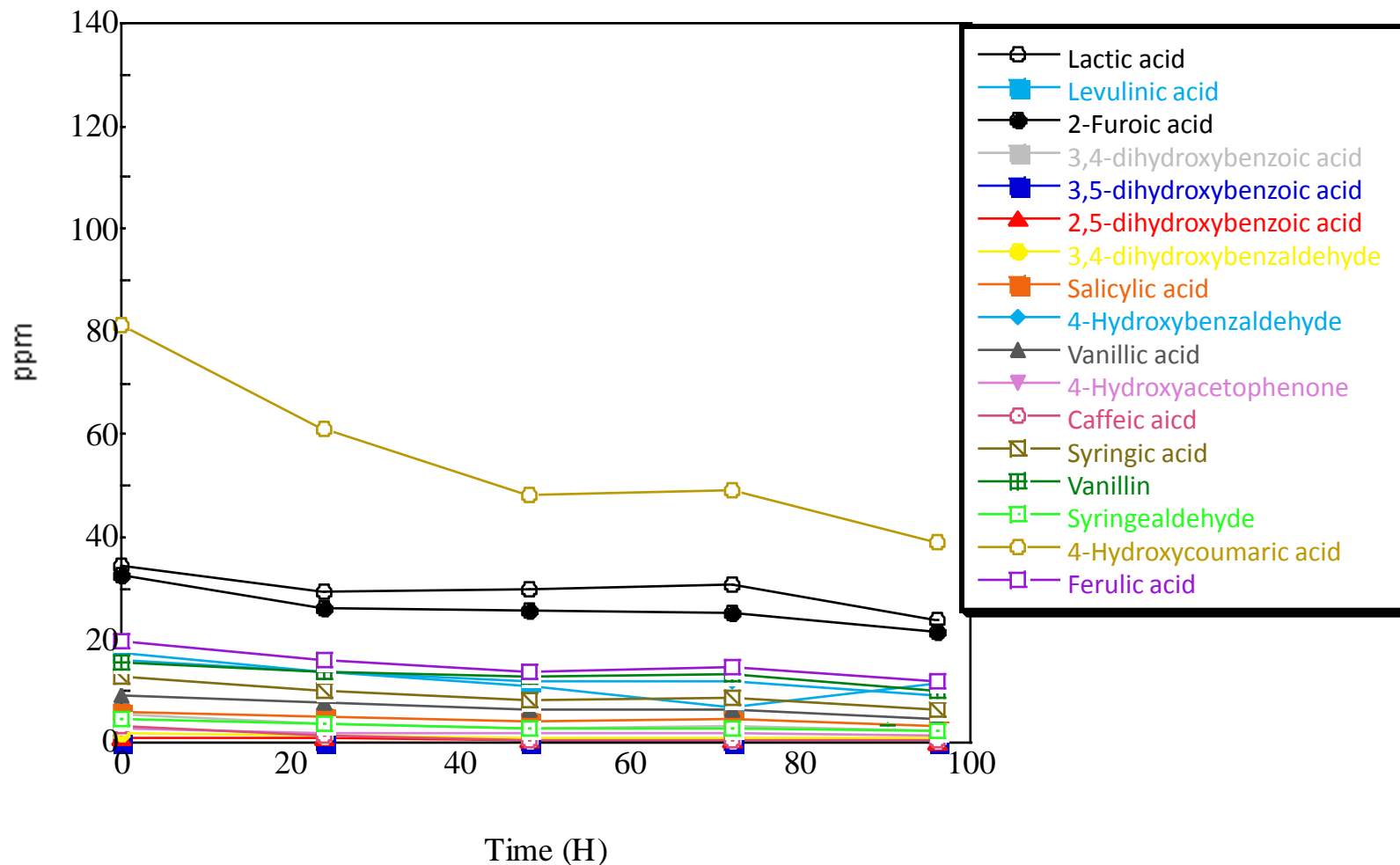
Time course of furfural & HMF metabolism



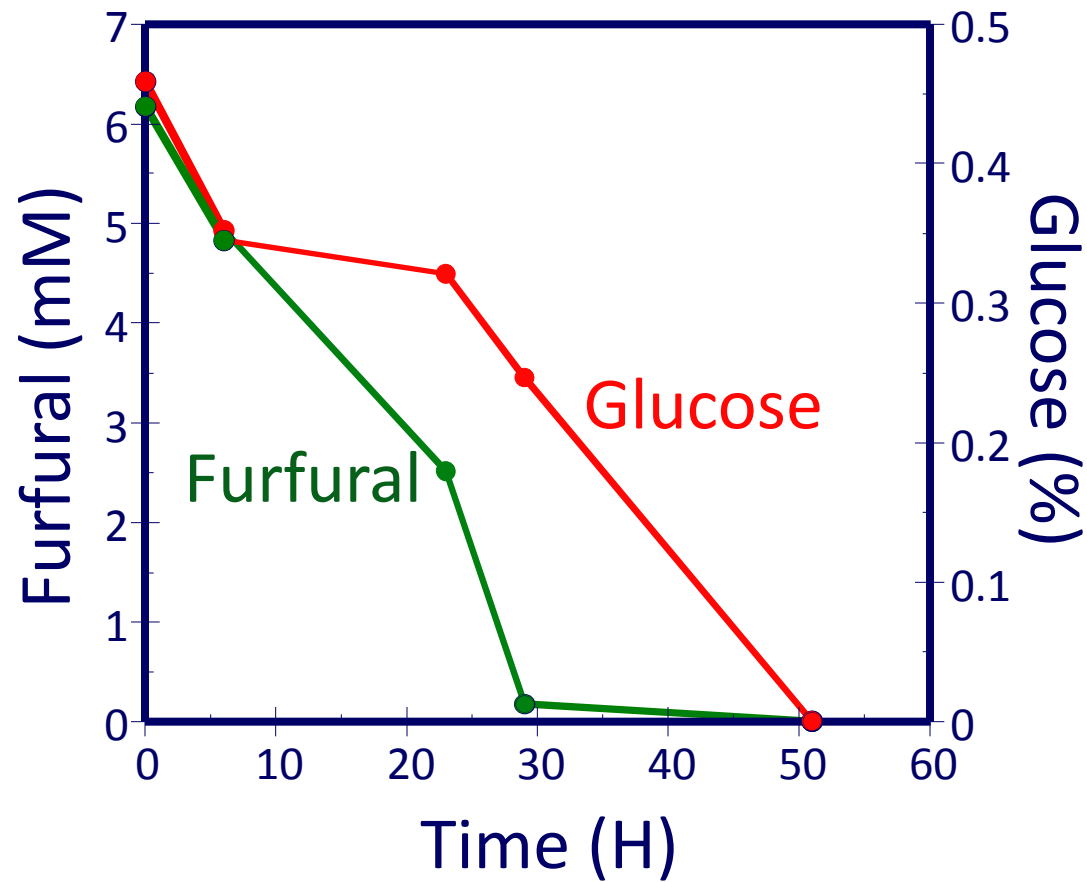
C8-treated hydrolysate



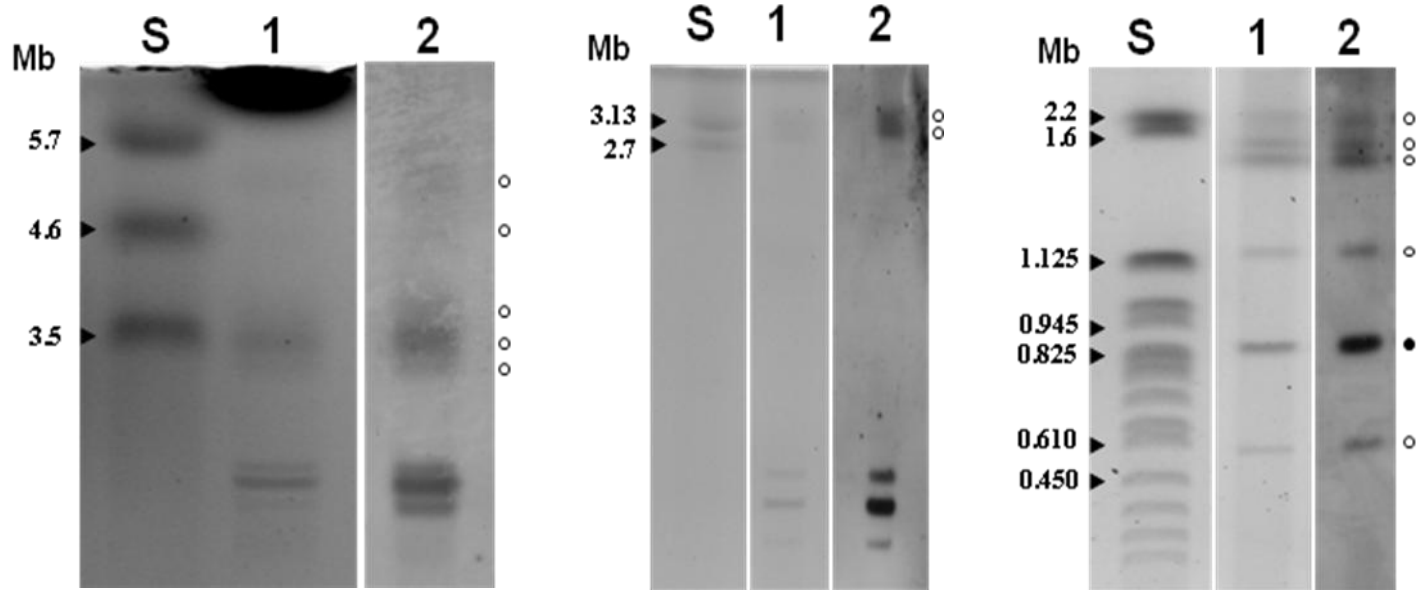
Untreated corn stover hydrolysate



Metabolism of furfural before glucose



Coniochaeta ligniaria genome



13 chromosomes (0.5-5.0 Mb)

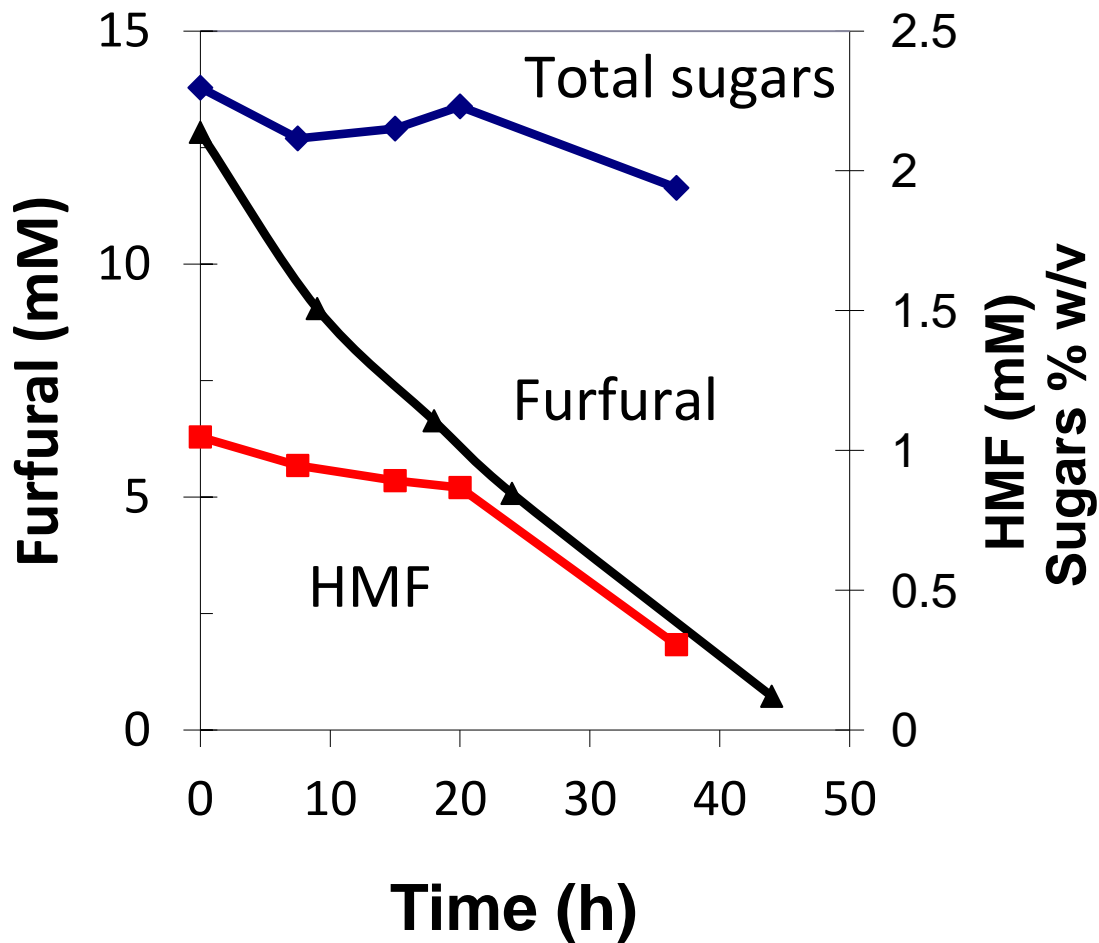
Estimated total size 30.1 Mb

2: (TTAGGG)₆

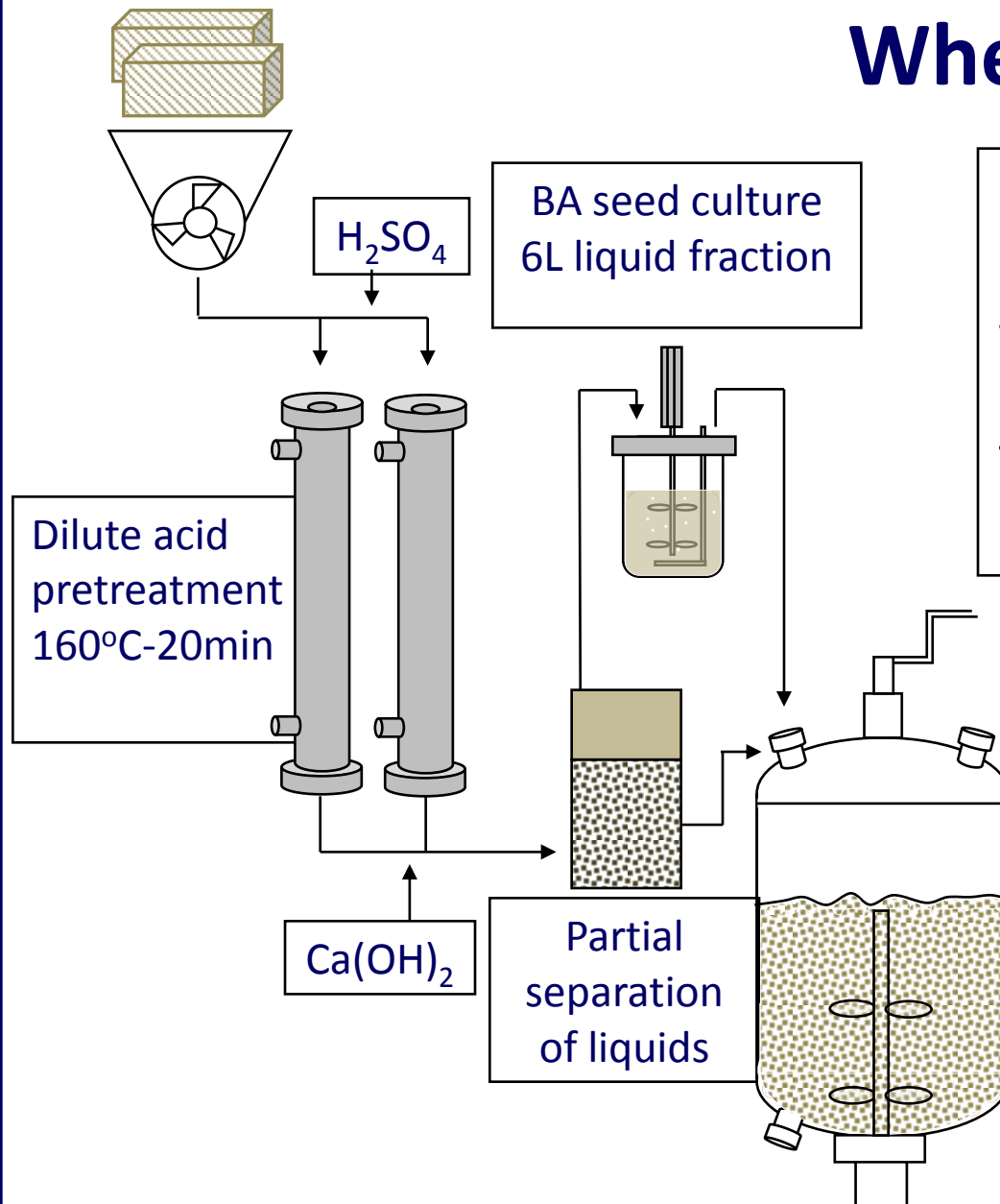
Characterizing *C. ligniaria*

- G + C content 49.2% (coding regions 51.9%)
- No unusual codons
- Haploid
- UV mutagenesis
- Transformation by protoplasting
- Antibiotic resistance: hygromycin, G418, benomyl

Wheat straw--bioabatement



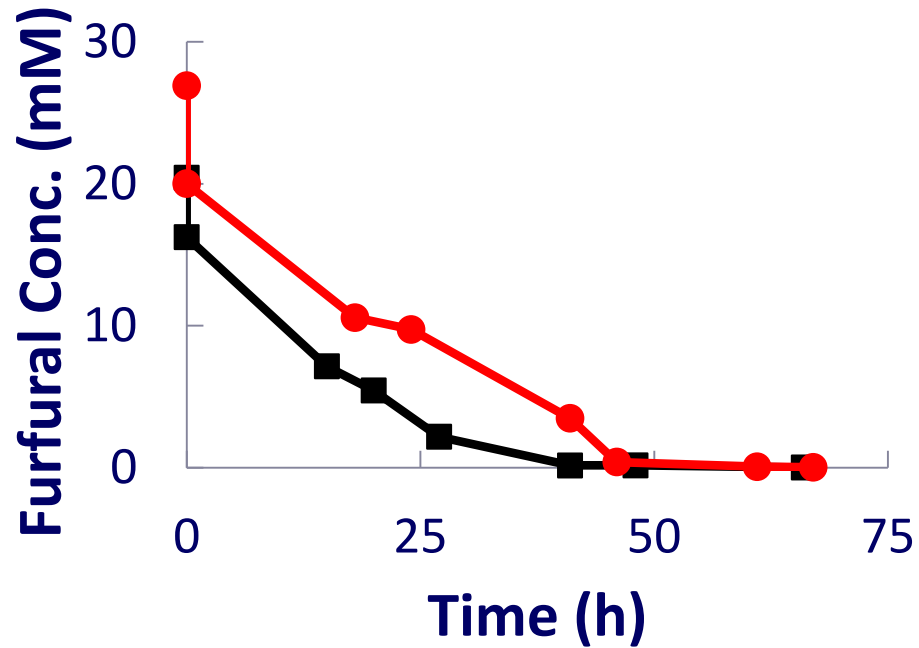
Wheat Straw SSF



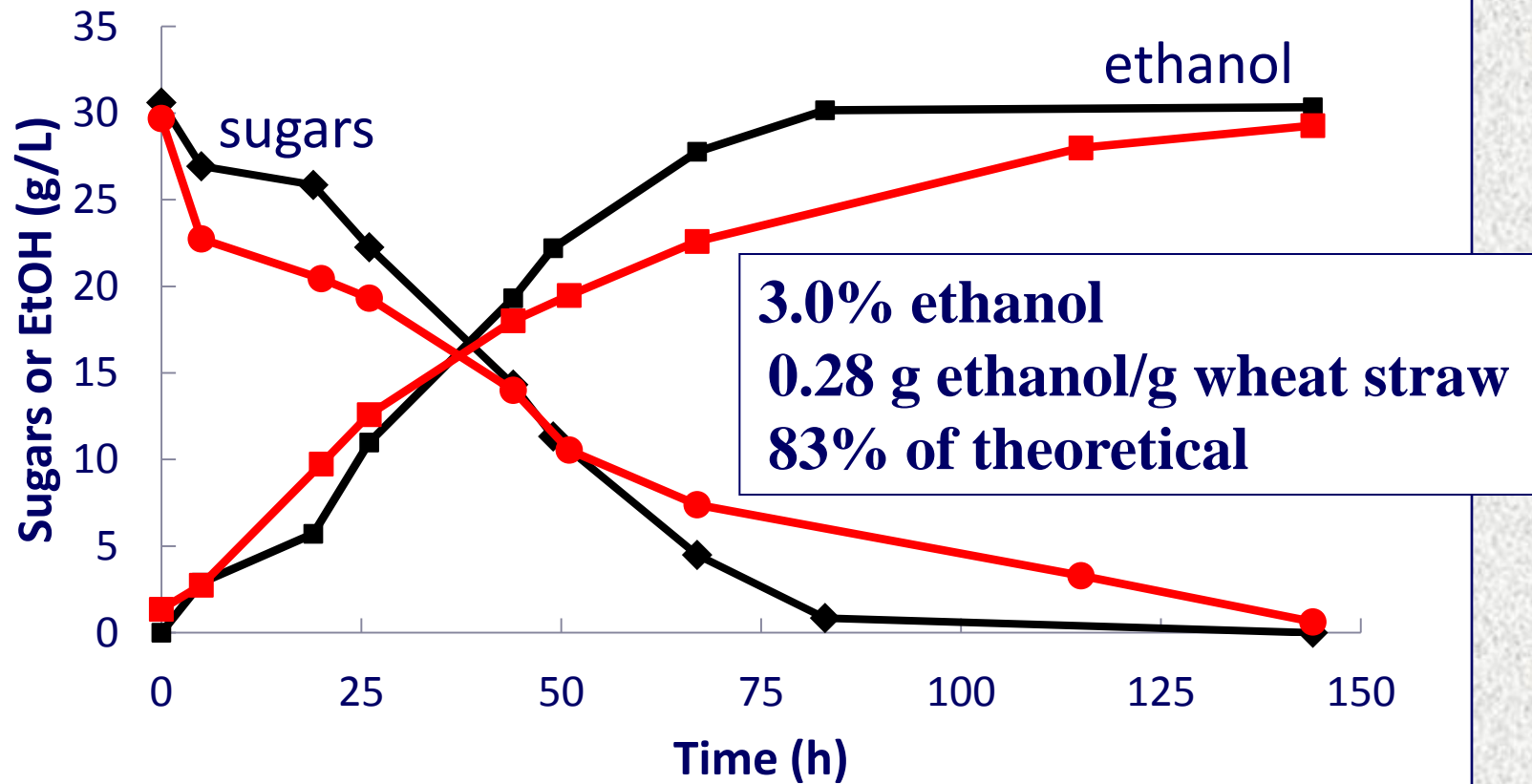
Multistep 60L SSF

1. NRRL30616 mitigation
2. *E. coli* pentose fermentation
3. Enzyme hydrolysis/hexose fermentation

Wheat straw bioabatement: furfural degradation



Wheat straw bioabatement: SSF



Summary

- ◆ A biological approach to inhibitor abatement
- ◆ Best isolate: Ascomycete *Coniochaeta ligniaria* C8
- ◆ Consumes furfural and HMF from hydrolyzed corn stover; also phenolics and organic acids
- ◆ Enhanced fermentation of biomass feedstocks: crop residues & energy crops

Acknowledgements

USDA/ARS/NCAUR

- ◆ Sarah Frazer, Greg Kennedy, Loren Iten

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